

This article was downloaded by:

On: 24 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Liquid Chromatography & Related Technologies

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713597273>

Multivariate Methods to Evaluate the Role of Mixed Supports in Reversed-Phase Thin-Layer Chromatography

Gábor Gullner^a; Tibor Cserhádi^b; Barna Bordás^b; Klára Valkó^b

^a Plant Protection Institute Hungarian Academy of Sciences, Budapest, Hungary ^b Central Research Institute for Chemistry Hungarian Academy of Sciences, Budapest, Hungary

To cite this Article Gullner, Gábor , Cserhádi, Tibor , Bordás, Barna and Valkó, Klára(1989) 'Multivariate Methods to Evaluate the Role of Mixed Supports in Reversed-Phase Thin-Layer Chromatography', *Journal of Liquid Chromatography & Related Technologies*, 12: 6, 957 – 978

To link to this Article: DOI: 10.1080/01483918908051773

URL: <http://dx.doi.org/10.1080/01483918908051773>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

MULTIVARIATE METHODS TO EVALUATE THE ROLE OF MIXED SUPPORTS IN REVERSED-PHASE THIN-LAYER CHROMATOGRAPHY

GÁBOR GULLNER¹, TIBOR CSERHÁTI²,
BARNA BORDÁS² AND KLÁRA VALKÓ²

¹*Plant Protection Institute
Hungarian Academy of Sciences
1022 Budapest*

Herman O. u. 15, Hungary
²*Central Research Institute for Chemistry
Hungarian Academy of Sciences
H-1025 Budapest
Pusztaszeri u. 59-67, Hungary*

ABSTRACT

Hydrophobic properties of 17 aniline and phenol derivatives were characterized by reversed-phase thin-layer chromatographic and high performance liquid chromatographic retention data.

In order to elucidate the role of thin-layer chromatographic supports in the hydrophobicity determination paraffin coated silica, aluminium oxide, cellulose, diatomaceous earth and their mixtures were used. Water, water-methanol 7:3 and 1 M NaCl served as mobile phases. The retention data were analyzed by spectral mapping technique.

The potency values differed from support to support proving that the composition of support has a deciding role in the hydrophobicity determination of aniline and phenol derivatives. The eluents did not influence considerably the potency order of supports.

The two dimensional non linear mapping of spectral data clearly showed that each sorbent influenced considerably the selectivity.

Good linear correlations were found between the reversed-phase high performance liquid chromatographic parameters of the compounds in partially dissociated state and the R values measured on supports containing high percentage of silica and diatomaceous earth.

INTRODUCTION

Hydrophobic properties of compounds are one of the most important physico-chemical parameters used in quantitative structure-activity relationship studies /1-3/. Lipophilic properties of compounds can be characterized by the logarithm of octanol-water partition coefficients (log P) measured by shake-flask method /4/. Correlation was found between reversed-phase thin-layer chromatographic (RP-TLC) retention data /5-7/, high performance liquid chromatographic (HPLC) retention data /8-10/ and gas-liquid chromatographic (GLC) retention data /11-13/ and log P values. All of these correlations depend on the similarity of the chromatographic partition systems to the octanol/water partition system used as reference.

Recent results indicate that in RP-TLC the adsorption characteristics of the impregnated supports also influence considerably the retention /14, 15/. In the case of 5-nitroimidazole derivatives it was shown that the quality of the impregnating agent influenced considerably the R_M values, however, the correlations

between the R_M values determined on silica plates impregnated with various lipophilic agents were very good /16/. These lipophilicity values have been successfully correlated with the biological activity data /17/.

It was also revealed that for polar compounds not only the adsorptive strength but also the surface pH values of supports has a considerable influence on the lipophilicity determination /18/.

The aim of our present study was to evaluate the role of thin-layer supports in the traditional RP-TLC retention behaviour of basic (aniline) and acidic (phenol) compounds. Silica and aluminium oxide were applied as strong, cellulose and diatomaceous earth as weak sorbents. Silica represented a sorbent with acidic, aluminium oxide with alkaline surface pH values. Their mixtures served as to show the continuity of the adsorptive properties of them. The new covalently bonded silica supports were omitted from our investigations, as their applications were reviewed in details by Brinkman and De Vries /19/.

Reversed-phase high performance liquid chromatographic retention data of the 17 model compounds were used as a reference measure of their lipophilicity.

EXPERIMENTAL

The following chromatographic sorbents were applied: Kieselgel 60 G (Merck Inc., Darmstadt, F. R.

G.), MN Aluminium oxide G (Macherey - Nagel Inc., Düren, F. R. G.), Cellulosepulver MN 300 (Macherey-Nagel) and Kieselgur G (Merck). The compositions of mixed supports used for the experiments are listed in Table 1.

The supports were mixed in the listed compositions and then a solution of 5% paraffin oil in n-hexane was added to each mixture to obtain support : paraffin oil 95:5 weight ratio. The suspensions were shaken overnight at room temperature then the n-hexane was evaporated in a rotary vacuum evaporator. Layers of 0.25 mm thickness were prepared on 20 x 20 cm glass plates from each sorbent.

Ortho-, meta-, and para-nitroaniline, 2,4-, 2,6-dinitroaniline, 2,4,6-trinitro-aniline, 2-chloro-4-nitroaniline, 4-chloro-3-nitroaniline, 2,6-dichloro-4-nitroaniline as well as para-nitrophenol, 2,4- and 2,6-dinitrophenol, 2,4,6-trinitrophenol, 3,5-dinitro-4-cyanophenol, 3-nitro-4-cyano-5-chlorophenol, 3-nitro-4-cyano-5-bromophenol and 3-nitro-4-cyano-5-iodophenol served as alkaline and acidic model compounds.

The compounds were dissolved in acetone at a concentration of 2 mg/ml, and 5 μ l of each solution was spotted on the plates.

Water /I/, water-methanol 7:3 (v/v) /II/ and 1 M NaCl /III/ were applied as mobile phases.

Table 1. Compositions of mixed supports.

| No. of support | Kieselgel % | Aluminium oxide % | Cellulose % | Kieselgur % |
|----------------|-------------|-------------------|-------------|-------------|
| 1 | 100 | - | - | - |
| 2 | - | - | 100 | - |
| 3 | - | 100 | - | - |
| 4 | - | - | - | 100 |
| 5 | 80 | - | 20 | - |
| 6 | 80 | 20 | - | - |
| 7 | 80 | - | - | 20 |
| 8 | 60 | - | 40 | - |
| 9 | 60 | 40 | - | - |
| 10 | 60 | - | - | 40 |
| 11 | 40 | - | 60 | - |
| 12 | 40 | 60 | - | - |
| 13 | 40 | - | - | 60 |
| 14 | 20 | - | 80 | - |
| 15 | 20 | 80 | - | - |
| 16 | 20 | - | - | 80 |
| 17 | - | 20 | 80 | - |
| 18 | - | - | 80 | 20 |
| 19 | - | 40 | 60 | - |
| 20 | - | - | 60 | 40 |
| 21 | - | 60 | 40 | - |
| 22 | - | - | 40 | 60 |
| 23 | - | 80 | 20 | - |
| 24 | - | - | 20 | 80 |
| 25 | - | 80 | - | 20 |
| 26 | - | 60 | - | 40 |
| 27 | - | 40 | - | 60 |
| 28 | - | 20 | - | 80 |
| 29 | 60 | 20 | 20 | - |
| 30 | 60 | - | 20 | 20 |
| 31 | 60 | 20 | - | 20 |
| 32 | 40 | 20 | 40 | - |
| 33 | 40 | - | 40 | 20 |
| 34 | 40 | 40 | - | 20 |
| 35 | 40 | 40 | 20 | - |
| 36 | 40 | - | 20 | 40 |
| 37 | 40 | 20 | - | 40 |
| 38 | 20 | 40 | 40 | - |
| 39 | 20 | 60 | 20 | - |
| 40 | 20 | 20 | 60 | - |
| 41 | 20 | - | 40 | 40 |
| 42 | 20 | - | 20 | 60 |
| 43 | 20 | - | 60 | 20 |
| 44 | 20 | 40 | - | 40 |
| 45 | 20 | 60 | - | 20 |

(continued)

Table 1. /Continued/

| No. of support | Kieselgel % | Aluminium oxide % | Cellulose % | Kieselgur % |
|----------------|-------------|-------------------|-------------|-------------|
| 46 | 20 | 20 | - | 60 |
| 47 | - | 20 | 60 | 20 |
| 48 | - | 40 | 40 | 20 |
| 49 | - | 20 | 40 | 40 |
| 50 | - | 40 | 20 | 40 |
| 51 | - | 20 | 20 | 60 |
| 52 | - | 60 | 20 | 20 |
| 53 | 40 | 20 | 20 | 20 |
| 54 | 20 | 20 | 40 | 20 |
| 55 | 20 | 40 | 20 | 20 |
| 56 | 20 | 20 | 20 | 40 |

After development, the compounds were detected visually. For each experiment five independent parallel determinations were carried out.

The measured retention data were analyzed separately for each eluent by spectral mapping technique (20) in order to separate the retention strength (potency =P) and selectivity of supports. To facilitate calculations the behaviour of compounds was characterized by $100 * R_f$ instead of R_f value. This method has been successfully adapted to classify eluents /21/ and covalently bonded sorbents /22/ in RP-TLC. The two dimensional non-linear mapping of selectivity spectra was carried out by the method published in ref. 23.

To assess the role of the eluents in the selectivity in RP-TLC separation, linear correlations

were calculated between the potency orders determined for each eluent. The significance of the differences between the r values was checked by the z -test. To evaluate the individual contribution of pure supports to the overall retention multivariate linear regressions were calculated between the separate potency values (dependent variables) and the data of Table 1 (independent variables) according to the equation bellow:

$$y = a + b_1 x_1 + b_2 x_2 + b_3 x_3 + b_4 x_4$$

where y = potency values of eluents I, II, and III;

x_1 = ratio of Kieselgel in the mixed support %/,

x_2 = ratio of aluminium oxide in the mixed support %/,

x_3 = ratio of cellulose in the mixed support %/,

x_4 = ratio of Kieselgur in the mixed support %/

The RP-HPLC system consisted of a Liquopump Model 312 (LaborMIM, Budapest, Hungary) with modified electronics and Altex (Cotati, Calif., U.S.A.) check valves, a Rheodyne (Cotati, Calif., U.S.A.) Model 7010 sample injection valve with 20 μ l loop, a 250 x 4.6 mm column with Dimesil C-18 (10 μ m) (Chromatronix Inc., Calif., U. S. A.) stationary phase and an ISCO (Lincoln, Nebraska, U. S. A.) Model 226 absorbance monitor with 254 nm source screen and an Endim Model 621.01 (Berlin, GDR) recorder. Retention time measurements and data processing were carried out on an Apple II.

microcomputer (Apple Computer Inc., Cupertino, Calif., U.S.A.).

To determine the $\log k'$ values of the undissociated compounds the eluents were mixtures of methanol and 0.025 M phosphate buffers, pH = 1.70 for phenols and pH = 7.99 for the anilines, ranging from 65 to 80 % (v/v) methanol (RP-HPLC system A). The $\log k'$ values were also determined using the same pH for both type of compounds, i. e. 0.025 M phosphate buffer pH = 4.35 (RP-HPLC system B). In both cases the flow rate was 1.00 ml/min; 0.1 mg of the compound tested and 2 mg NaNO_3 were dissolved in 5 ml eluent and 20 μl of the solutions was injected onto the column. The retention time of NaNO_3 was regarded as the dead time (t_0). Both retention time (t_R) and the dead time were measured three times. The average $\log k'$ values were calculated. The $\log k'$ values obtained using eluents with various methanol concentrations were plotted against the methanol concentration. As both the intercepts ($\log k'_0$) and the slopes (b) of the obtained straight lines may correlate to the $\log P$ values /24, 25/, linear correlations were separately calculated between the 4 types of RP-HPLC parameters ($\log k'_0$, b referring to systems A, and B, respectively) and the 168 series of R_M values determined on 56 support mixtures using 3 types of eluent system.

RESULTS AND DISCUSSION

The potency values calculated from the $100 \times R_f$ values are compiled in Table 2. These values show the retention strengths on the given support. The considerable difference between the retention strength of support impregnated exactly the same way proved again that the supports retain their original adsorptive characteristics that is the R_f values of compounds depend considerably on the type of support.

Table 2. The potency (P values) calculated from R_f values for the various mixed supports.

| No. of support | Type of the eluent | | |
|----------------|--------------------|-----|-----|
| | I | II | III |
| 1 | 144 | 298 | 94 |
| 2 | 232 | 307 | 160 |
| 3 | 54 | 252 | 48 |
| 4 | 388 | 412 | 388 |
| 5 | 158 | 279 | 103 |
| 6 | 136 | 272 | 83 |
| 7 | 172 | 310 | 111 |
| 8 | 162 | 266 | 105 |
| 9 | 121 | 254 | 78 |
| 10 | 171 | 307 | 106 |
| 11 | 161 | 261 | 109 |
| 12 | 93 | 243 | 76 |
| 13 | 193 | 320 | 140 |
| 14 | 165 | 264 | 122 |
| 15 | 73 | 223 | 67 |
| 16 | 221 | 348 | 170 |
| 17 | 157 | 237 | 128 |
| 18 | 256 | 300 | 191 |
| 19 | 131 | 231 | 110 |
| 20 | 268 | 310 | 215 |
| 21 | 117 | 214 | 106 |
| 22 | 296 | 326 | 230 |

(continued)

Table 2. /Continued/

| No. of support | Type of the eluent | | |
|----------------|--------------------|-----|-----|
| | I | II | III |
| 23 | 114 | 217 | 97 |
| 24 | 321 | 351 | 288 |
| 25 | 99 | 258 | 82 |
| 26 | 135 | 298 | 114 |
| 27 | 166 | 329 | 162 |
| 28 | 241 | 355 | 209 |
| 29 | 148 | 255 | 103 |
| 30 | 176 | 289 | 123 |
| 31 | 153 | 281 | 97 |
| 32 | 155 | 262 | 110 |
| 33 | 177 | 272 | 125 |
| 34 | 138 | 281 | 91 |
| 35 | 125 | 240 | 98 |
| 36 | 179 | 286 | 134 |
| 37 | 152 | 289 | 104 |
| 38 | 128 | 244 | 100 |
| 39 | 113 | 243 | 92 |
| 40 | 160 | 255 | 122 |
| 41 | 195 | 289 | 150 |
| 42 | 204 | 307 | 156 |
| 43 | 172 | 276 | 129 |
| 44 | 135 | 303 | 108 |
| 45 | 99 | 258 | 75 |
| 46 | 186 | 316 | 150 |
| 47 | 193 | 277 | 146 |
| 48 | 155 | 251 | 134 |
| 49 | 187 | 285 | 169 |
| 50 | 131 | 254 | 114 |
| 51 | 205 | 313 | 178 |
| 52 | 128 | 255 | 114 |
| 53 | 154 | 277 | 113 |
| 54 | 156 | 260 | 114 |
| 55 | 128 | 259 | 98 |
| 56 | 169 | 290 | 134 |

The P values obtained for the three types of mobile phases correlated with each other fairly well as it is shown by equ. 1 - 3.

$$P_{II} = 192.9 (\pm 8.8) + 0.525 (\pm 0.050)P_I \quad (1)$$

$$n = 56 \quad r = 0.821$$

$$P_{III} = -20.9 (\pm 6.9) + 0.903(\pm 0.039)P_I \quad (2)$$

$$n=56 \quad r=0.953$$

$$P_{III} = -198.8 (\pm 35.1) + 1.172 (\pm 0.124)P_{II} \quad (3)$$

$$n=56 \quad r=0.790$$

Equations 1-3 suggests that the eluent composition did not influence considerably the retention order of the compounds investigated on the mixed supports. Eluent II was stronger than eluent I, according to the general rule, however eluent III exposed a lower eluent strength than eluent I. This observation was somewhat unexpected because the ions present in the eluents are bond to the available silanol groups non covered by paraffin oil. This phenomenon could have reduced the retention capacity of supports. However, an increase of retention was found in the presence of ions. It is assumed that the ions suppress the dissociation of polar amino and hydroxyl groups increasing in this way the R_M values of compounds /26/.

The z -test showed that the correlation coefficient of $P_{III} - P_I$ relationship was significantly higher than the other two correlation coefficients (significance level was over 99.9%), but between the fitness of $P_{II} - P_I$ and $P_{III} - P_{II}$ relationships no significant difference was found ($t=0.46$). This finding suggests that eluent pairs I - III showed a higher similarity than the eluent pairs I - II and II - III.

The potency values of supports did not show linear correlations to the percentage of their compositions (F values were 1.45, 0.75 and 2.89 for eluent I, II and III, respectively). The lack of correlations can be explained in two ways, namely, either the correlation is non linear or the supports interact with each other. The two dimensional non linear maps of selectivity data with water as eluent (Fig. 1.) shows, that the support mixtures containing silica or alumina form distinct clusters. It means that the supports retain their original selectivity character also after impregnation. Of course, the support mixtures containing the both sorbents overlap in the map.

The situation is similar for the other two eluents (Fig. 2 and 3.). The cluster formation prevails independently of the eluent composition. These observations indicate that the composition of support has a predominant role in the selectivity even in RP-TLC separation of polar compounds and the eluent composition is of minor importance.

The parameters of the linear relationships between the $\log k'$ values and the methanol concentrations of the eluent in the RP-HPLC measurements are shown in Table 3. The correlation coefficients were over 99% significance level in each case. The values of $\log k'_0$ and the slope (b) of the aniline derivatives are the

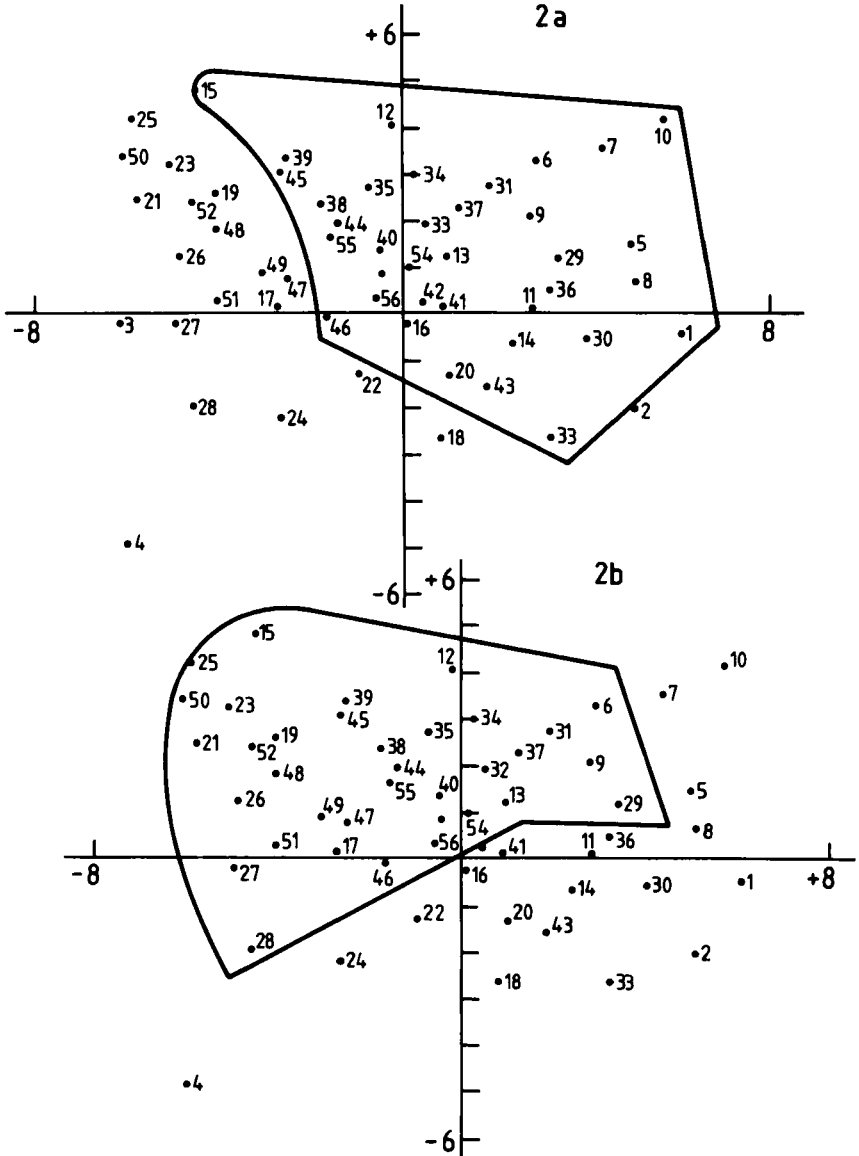


Figure 2.
Two dimensional non linear maps of selectivity data obtained by using water - methanol (7:3) as eluent. Number of iterations: 8. Error of mapping: 0.025. Numbers indicates supports listed in Table 1.
a: supports containing silica encircled
b: supports containing aluminium oxide encircled

Downloaded At: 13:55 24 January 2011

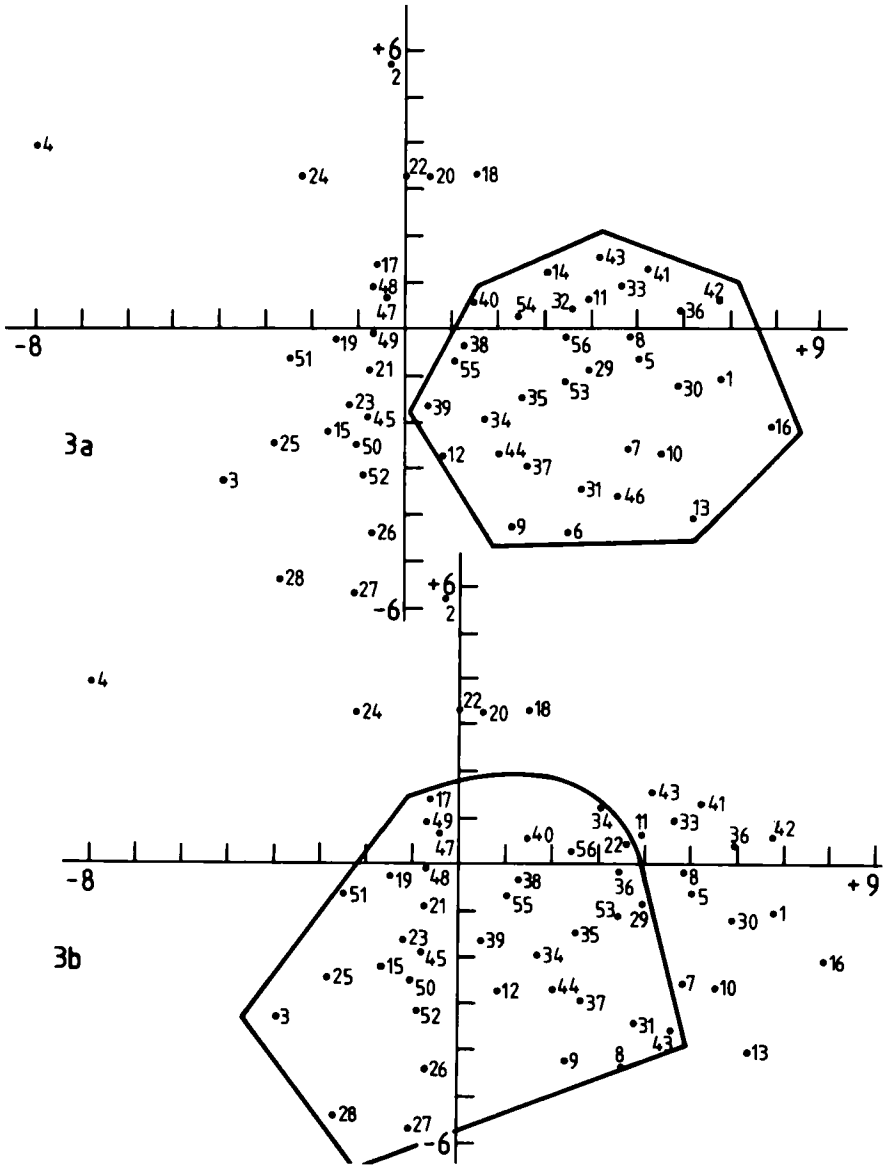


Figure 3.

Two dimensional non linear maps of selectivity data obtained by using 1M NaCl as eluent. Number of iterations: 10. Error of mapping: 0.0246. Numbers indicate supports listed in Table 1.

a: supports containing silica encircled

b: supports containing aluminium oxide encircled

Table 3.

Parameters of the linear relationships between the $\log k'$ values and the methanol concentration of the eluent (C%) determined by RP-HPLC. ($\log k' = \log k'_0 + bC\%$)

| Compounds | * System A (undissociated form) | | * System B (partially dissociated form) | |
|--|---------------------------------------|-------------|--|-------------|
| | 100b | $\log k'_0$ | 100b | $\log k'_0$ |
| o-NO ₂ -aniline | -2.48 | 1.507 | -2.48 | 1.507 |
| m-NO ₂ -aniline | -2.37 | 1.253 | -2.37 | 1.253 |
| p-NO ₂ -aniline | -2.14 | 0.980 | -2.14 | 0.980 |
| 2,4-(NO ₂) ₂ -aniline | -2.67 | 1.715 | -2.67 | 1.715 |
| 2,4,6-(NO ₂) ₃ -aniline | -2.64 | 1.676 | -2.64 | 1.676 |
| 2-Cl-4-NO ₂ -aniline | -2.80 | 1.879 | -2.80 | 1.879 |
| 4-Cl-3-NO ₂ -aniline | -2.89 | 1.814 | -2.89 | 1.814 |
| 2,6-(Cl) ₂ -4-NO ₂ -an. | -3.29 | 2.568 | -3.29 | 2.568 |
| p-NO ₂ -phenol | -2.58 | 1.486 | -8.83 | 5.044 |
| 2,4-(NO ₂) ₂ -phenol | -2.90 | 1.789 | -12.24 | 5.699 |
| 2,6-(NO ₂) ₂ -phenol | -5.17 | 2.984 | -8.85 | 3.649 |
| 2,4,6-(NO ₂) ₃ -phenol | -4.90 | 2.487 | -12.10 | 6.325 |
| 3,5(NO ₂) ₂ 4CN-phenol | -4.21 | 2.417 | -11.60 | 5.305 |
| 3NO ₂ , 4CN, 5Cl-phenol | -3.25 | 2.332 | -9.96 | 4.620 |
| 3NO ₂ , 4CN, 5Br-phenol | -4.24 | 2.684 | -12.80 | 7.009 |
| 3NO ₂ , 4CN, 5I-phenol | -2.85 | 1.592 | -11.80 | 5.452 |

*

see Experimental

same using the two different eluent systems that is the dissociation state of these compounds does not change in this pH region significantly (from pH=4.35 to pH=7.99), resulting in unchanged chromatographic retention.

The linear relation of the R_M values to the RP-HPLC retention parameters ($\log k'_0$ and b) was characterized by correlation coefficient. The correlation coefficients were calculated for the 3 times 56 R_M values and for the 4 types of RP-HPLC retention data, and their significance level was also investigated. The results are summarized in Table 4.

The results show that higher correlation coefficients were obtained when R_M values were correlated to the RP-HPLC retention data obtained in eluent system B in which the pH was 4.35 and the compounds were partially dissociated form. As the R_M values were obtained without using buffer in that case the compounds were also partially dissociated form.

It is also noticeable that R_M values obtained by using eluent III (i.e. 1 M NaCl) do not show as many high correlation coefficients with the RP-HPLC retention data obtained with mobile phase B as in case of the other two eluent systems. Salts in the mobile phase can modify the retention behaviour of polarizable compounds in several ways. They can adsorb on the free silanol groups of silica surface not covered by paraffin oil, they can lessen the degree of dissociation of polar

Table 4.

The significance of the correlation coefficients (r) for the relation of 4 RP-HPLC paramers to the 168 series of R_M values (obtained on 56 support mixtures using 3 eluent systems).

| RP-HPLC data | RP-TLC R_M obtained by | Significance level of r (%) | | | |
|-----------------|--------------------------|-------------------------------|-------|---------|-----------|
| | | under 98 | 98-99 | 99-99.9 | over 99.9 |
| | | Number of r values | | | |
| $\log k'_o$ (A) | eluent I | 37 | 5 | 13 | 1 |
| b (A) | eluent I | 52 | - | 4 | - |
| $\log k'_o$ (B) | eluent I | 12 | 2 | 7 | 35 |
| b (B) | eluent I | 11 | 3 | 7 | 35 |
| $\log k'_o$ (A) | eluent II | 42 | 5 | 8 | 1 |
| b (A) | eluent II | 54 | 2 | - | - |
| $\log k'_o$ (B) | eluent II | 8 | 2 | 8 | 38 |
| b (B) | eluent II | 12 | 4 | 8 | 32 |
| $\log k'_o$ (A) | eluent III | 51 | 4 | 1 | - |
| b (A) | eluent III | 56 | - | - | - |
| $\log k'_o$ (B) | eluent III | 17 | 2 | 13 | 24 |
| b (B) | eluent III | 20 | 2 | 23 | 11 |

compounds, they increase the dielectric constant and surface tension of water, which also influence the retention and they can modify the intramolecular forces between polar substituents. The influence of salts on the retention of polar compounds depends on the processes mentioned above making unpredictable their

Table 5.

Parameters of the five best correlations between the 4 RP-HPLC retention data (Y) and the R_M values (X). (r stands for the correlation coefficient, s is the standard deviation.)

$$Y = a + b \cdot X$$

| RP-HPLC data (Y) | RP-TLC data(X) | | Parameters of correlation | | | |
|---------------------|----------------|--------|---------------------------|-------|-------|-------|
| | Support | Eluent | a | b | s | r |
| $\log k'_0$ (A) | 18 | I | 1.62 | -0.68 | 0.196 | 0.681 |
| $\log k'_0$ (A) | 22 | I | 1.45 | -0.66 | 0.201 | 0.661 |
| $\log k'_0$ (A) | 20 | I | 1.58 | -0.67 | 0.223 | 0.629 |
| $\log k'_0$ (A) | 2 | I | 1.72 | -0.74 | 0.250 | 0.625 |
| $\log k'_0$ (A) | 2 | II | 1.65 | -0.16 | 0.059 | 0.583 |
| b (A) | 18 | I | -0.026 | 0.013 | 0.003 | 0.753 |
| b (A) | 22 | I | -0.023 | 0.012 | 0.003 | 0.722 |
| b (A) | 2 | I | -0.028 | 0.015 | 0.004 | 0.718 |
| b (A) | 2 | II | -0.026 | 0.003 | 0.001 | 0.709 |
| b (A) | 20 | I | -0.026 | 0.013 | 0.003 | 0.704 |
| $\log k'_0$ (B) | 22 | I | 1.32 | -3.33 | 0.469 | 0.885 |
| $\log k'_0$ (B) | 5 | I | 4.24 | -2.55 | 0.367 | 0.881 |
| $\log k'_0$ (B) | 30 | I | 3.92 | -2.87 | 0.428 | 0.874 |
| $\log k'_0$ (B) | 42 | I | 3.37 | -3.29 | 0.493 | 0.872 |
| $\log k'_0$ (B) | 5 | II | 2.09 | -2.69 | 0.390 | 0.872 |
| b (B) | 7 | II | -0.028 | 0.021 | 0.002 | 0.946 |
| b (B) | 22 | I | -0.016 | 0.078 | 0.007 | 0.943 |
| b (B) | 1 | II | -0.031 | 0.021 | 0.002 | 0.938 |
| b (B) | 30 | II | -0.029 | 0.072 | 0.007 | 0.935 |
| b (B) | 5 | II | -0.035 | 0.062 | 0.006 | 0.932 |

effects and decreasing the correlation between the R_m values and RP-HPLC parameters.

From Table 4. it also can be seen, that about the same number of good correlation was found when $\log k'_0$ or the b values were used. It means that the slope and the intercept values correlate with each other.

The parameters of the 5 best correlations for each RP-HPLC parameter are compiled in Table 5. It is noticeable that the best correlations were obtained in the case of supports containing high percentage of silica and diatomaceous earth.

These results proved that the multivariate methods are suitable to elucidate adequately the role of supports in RP-TLC and they promote the better understanding of physico-chemical parameters influencing the retention in RP-TLC.

REFERENCES

1. T. Fujita, J. Iwasa and C. Hansch, *J. Am. Chem. Soc.* 86 (1964) 5175-5183.
2. C. Hansch and J. M. Clayton, *J. Pharm. Sci.*, 62 (1973) 1-13.
3. C. Hansch and W. J. Dunn, *J. Pharm. Sci.*, 61 (1972) 1-9.
4. C. Hansch and S. M. Anderson, *J. Org. Chem.*, 32 (1967) 2583-2592.
5. C. B. C. Boyce and B. V. Milborrow, *Nature*, 208 (1965) 537-538.
6. G. L. Biagi, A. M. Barbaro and M. C. Guerra, *J. Chromatogr.*, 41 (1969) 371-383.

7. G. L. Biagi, M. C. Guerra, A. M. Barbaro and M. F. Gamba, *J. Med. Chem.* 13 (1970) 511-518.
8. W. J. Haggert and E. A. Murrill, *Res. Develop.*, 25 (1974) 30-38.
9. J. M. McCall, *J. Med. Chem.*, 18 (1975) 549-563.
10. M. S. Mirlees, S. J. Moulton, C. T. Murphy and P. J. Taylor, *J. Med. Chem.*, 19 (1976) 615-623.
11. D. R. Clifford and D. A. M. Watkins, *Pestic. Sci.*, 2 (1971) 41-53.
12. E. János, B. Bordás and T. Cserhádi, *J. Chromatogr.*, 286 (1984) 63-67.
13. K. Valkó and A. Lopata, *J. Chromatogr.* 252 (1982) 77-90.
14. W. V. van Giesen and L. H. M. Janssen, *J. Chromatogr.*, 237 (1982) 199-211.
15. T. Cserhádi, *Chromatographia*, 18 (1984) 18-20.
16. M. C. Guerra, A. M. Barbaro, G. Cantelli Forti, M. T. Foffani, G. L. Biagi, P. A. Borea and A. Fini, *J. Chromatogr.*, 216 (1981) 93-102.
17. G. L. Biagi, A. M. Barbaro, M. C. Guerra, G. Cantelli Forti, G. Aicardi and P. A. Borea, *Teratogen. Carcin. Mut.*, 3 (1983) 429-438.
18. T. Cserhádi, Y. M. Darwin and Gy. Matolcsy, *J. Chromatogr.*, 270 (1983) 97-104.
19. U. A. Th. Brinkman and G. De Vries, *Proceedings of the Second International Symposium on Instrumental High Performance Thin-Layer Chromatography, Interlaken, 1982.* pp. 11-24.
20. P. J. Lewi, *Arzneim.-Forsch.*, 26 (1976) 1295-1303.
21. T. Cserhádi and B. Bordás, *J. Chromatogr.*, 286 (1984) 131-137.
22. T. Cserhádi, B. Bordás, L. Ekiert and J. Bojarski, *J. Chromatogr.*, 287 (1984) 385-390.
23. J. W. Jr. Sammon, *IEEE Trans. Comput.* C18 (1969) 411-415.

24. K. Valkó, J. Liquid Chromatogr., 7 (1984) 1405-1424.
25. T.Cserhádi, M. Szögyi and L. Györfi, Chromatographia 20 (1985) 253-260.
26. J. Kojima and S.S. Davis, Internat. J. Pharmaceut., 20 (1984) 203-207.